

BIOSYNTHESIS OF THE SESQUITERPENOID PHYTOALEXIN RISHITIN FROM ACETATE VIA OXYLUBIMIN IN POTATO

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(Received 28 March 1978)

Key Word Index—*Solanum tuberosum* × *S. demissum*; Solanaceae; potato; sesquiterpenoid phytoalexins; biosynthesis; rishitin; oxylubimin.

INTRODUCTION

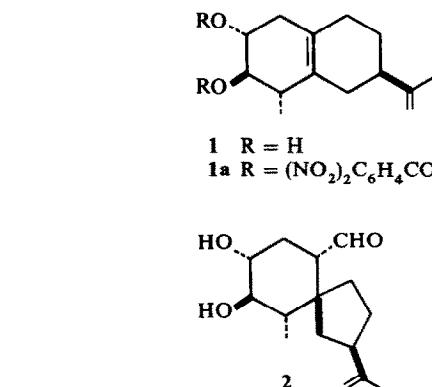
Since the isolation of rishitin (1) [1-3], many sesquiterpenoid phytoalexins and related compounds have been isolated from infected potatoes [3, 4]. Recently, attempts have been made to relate these compounds biogenetically. Stoessl *et al.* [4] proposed a pathway in which rishitin (1) and oxylubimin (2) [5, 6] were synthesized from farnesyl pyrophosphate via *trans*- and *cis*-eudesmanes, respectively. On the other hand, Kalan and Osman [7] proposed that 1 is synthesized through solavetivone, isolubimin, lubimin and 2. However, there is no experimental evidence so far, which supports these hypothetical pathways. We report herein experimental evidence, indicating that oxylubimin (2) is converted to rishitin (1) and that the transformation from the spirovetivanes is a major pathway in the biosynthesis of the noreudesmane (1).

RESULTS AND DISCUSSION

The specific radioactivity of crystalline rishitin *bis* (3,5-dinitrobenzoate) (1a) is shown in Table 1. The radioactive 1 was obtained by addition of authentic cold 1 to

Table 1. Effect of recrystallization on the specific radioactivity of *bis*(3,5-dinitrobenzoate) of rishitin-¹⁴C extracted from the potato tissue treated with oxylubimin-¹⁴C

Recrystallization	Radioactivity (10 ⁻³ dpm)	Wt of <i>bis</i> (3,5-dinitrobenzoate) (mg)	Specific activity (10 ⁻³ dpm/mg)
1st	420	17.6	23.8
2nd	299	12.7	23.5
3rd	197	8.4	23.5



the rishitin fraction extracted from tuber disks, which were infected by an incompatible race of *Phytophthora infestans* and treated with ¹⁴C-labeled 2. The specific activity of 1a remained unchanged on repeated recrystallization. This result demonstrated that radioactivity of 2 is incorporated into 1, indicating that 2 is a precursor of 1 in the infected potato tuber tissue.

If 2 is an intermediate on a main biosynthetic pathway to 1, addition of cold 2 along with acetate-2-¹⁴C should result in a significant decrease in incorporation of radioactivity into 1. In both treatments 1 and 2 of experiment 2 (Table 2), acetate-2-¹⁴C was first added to the infected potato disks and then, in treatment 2, unlabeled 2 was added, while in treatment 1, none of 2 was added. Preliminary experiments showed that addition of unlabeled 2 did not reduce the rate of uptake of labeled acetate by the tuber disks, and also that addition of 0.25, 0.5 and 1 mg of labeled 2 to the acetate (15 µg) reduced incor-

Table 2. Incorporation of radioactivity of ¹⁴C labeled precursors into rishitin in potato tuber tissue

Treatment	Precursors		Yield (µg)	Radioactivity (10 ⁻³ dpm)	Specific activity (10 ⁻³ dpm/µg)
	Acetate	Oxylubimin (2)			
1	22 × 10 ⁶ dpm (15 µg)	—	59*	1322 (6%)†	22.4
2	22 × 10 ⁶ dpm (15 µg)	Cold oxylubimin (1 mg)	189*	177 (0.8%)†	0.94
3	Cold acetate (15 µg)	113 × 10 ⁴ dpm (1 mg)	189*	138 (12.2%)†	0.73 [1.55]‡

* Data from the experiment 3, the other data from experiment 2.

† % incorporation into rishitin.

‡ Dilution value.

poration of the radioactivity from the acetate into **1** by 20, 67 and 87%, respectively. Addition of unlabeled **2** (1 mg) resulted in a 7-fold decrease in the relevant incorporation of radioactivity of acetate-2-¹⁴C into **1**. In treatment 3 of experiment 2 (Table 2), ¹⁴C-labeled **2** was administered along with cold acetate to give 189 µg of **1** (measured by GLC analysis). Calculation from the radioactivity of produced **1** revealed that 122 µg of accumulated **1** resulted from **2** applied. These results established that oxylubimin (**2**) with a spirovetivane skeleton is an intermediate on the main biosynthetic pathway to rishitin (**1**) with a noreudesmane skeleton. In view of the currently accepted pathway in which spirovetivanes are produced from eudesmanes [8], the aforementioned result is noteworthy. The efficient incorporation and low dilution value (1.55) also exclude incorporation resulting from fragmentation of **2** followed by synthesis of **1** from some of the fragments.

EXPERIMENTAL

Oxylubimin-¹⁴C (2.8×10^6 dpm/mg) was prepared by treating potato tuber tissue with acetate-2-¹⁴C according to the method of ref. [9] for prep of rishitin-¹⁴C. Oxylubimin-¹⁴C was extracted with EtOAc from soln in the holes made in the tuber tissue of potato cv Rishiri (*Solanum tuberosum* \times *S. demissum*). The holes have previously been inoculated with zoospore suspension and then filled with acetate-2-¹⁴C. The extracts were separated by TLC over Si gel (Merck Si gel G plate, 0.5 mm thick) with C_6H_6 -EtOAc (2:3), and then purified by recrystallization from isopropyl ether. After sterilizing the surface with 0.1% NaClO, tubers of Rishiri were cut into disks, 1 mm thick, 16 mm dia, washed with streaming H_2O and then incubated at 18° for 20 hr. Zoospore suspension of *P. infestans*, race 0 containing antibiotic ceporan (4 mg/ml) was inoculated onto both surfaces of the disks. Inoculated disks were incubated at 18°.

Experiment 1. 20 hr after inoculation, 76 disks were placed in 4 ml of sterile H_2O containing 1 mg oxylubimin-¹⁴C, ceporan (4 mg/ml) and 10% aq. Me_2CO , and incubated at 18° for 24 hr. Rishitin was isolated from the incubated sample with $MeOH$ followed by separation by TLC and was crystallized as the bis(3,5-dinitrobenzoate) according to the method of ref. [10]. After recrystallization, the radioactivity was determined in the usual way. The recrystallization and radioactivity counting were repeated.

Experiment 2. 25 hr after inoculation, 11 disks (ca 2.2 g) were placed in 1 ml of sterile H_2O containing, in treatment 1, acetate-2-¹⁴C (10 µCi/15 µg), in treatment 3, 1 mg oxylubimin (1.13×10^6 dpm/mg) and cold NaOAc (15 µg), and then incubated at 18° for 12 hr. Each soln for the treatment contained 4 mg ceporan and 10% aq. OAc. Rishitin-¹⁴C was extracted and crystallized as described above.

Experiment 3. The coordinated experiment with experiment 2 was carried out. The disks were treated with unlabeled OAc or unlabeled oxylubimin as in experiment 2, and the content of rishitin was determined by GLC using a modified method of ref. [11], after separating the rishitin fraction by the method described above. Gaschromatograph (JEOL 1100, Nihondenshi Co.) fitted with a 300 \times 0.03 cm i-d, pyrex glass column packed with 3% silicone OV-225 on chromosorb W (80–100 mesh) was used. Operating conditions were: temp., column 190°, injector 220°, detector 215°: carrier gas N_2 45 ml/min: sample size 1 µl.

Acknowledgements—We wish to thank Messrs Y. Nishibe, Y. Umemura, Hokkaido Agric. Exp. Sta. for their generous supply of experimental materials.

REFERENCES

1. Tomiyama, K., Sakuma, T., Ishizaka, N., Katsui, N., Takasugi, M., and Masamune, T. (1968) *Phytopathology* **58**, 115.
2. Katsui, N., Murai, A., Takasugi, M., Imaizumi, K., Masamune, T. and Tomiyama, K. (1968) *J. Chem. Soc. Chem. Commun.* 43.
3. Masamune, T., Murai, A., Takasugi, M., Matsunaga, A., Katsui, N., Sato, N. and Tomiyama, K. (1977) *Bull. Chem. Soc. Jpn* **50**, 1201.
4. Stoessl, A., Stothers, J. B. and Ward, W. B. (1976) *Phytochemistry* **15**, 855.
5. Katsui, N., Matsunaga, A. and Masamune, T. (1974) *Tetrahedron Letters*, 4483; Katsui, N., Matsunaga, A., Kitahara, H., Yagihashi, F., Murai, A., Masamune, T. and Sato, N. (1977) *Bull. Chem. Soc. Jpn* **50**, 1217.
6. Stoessl, A., Stothers, J. B. and Ward, E. W. B. (1975) *J. Chem. Soc. Chem. Commun* 431
7. Kalan, E. B. and Osman, S. F. (1976) *Phytochemistry* **15**, 775.
8. Rücker, G. (1973) *Angew. Chem. Int. Ed. Engl.* **12**, 793.
9. Horikawa, T., Tomiyama, K. and Doke, N. (1976) *Phytopathology* **66**, 1186.
10. Ishiguri, Y., Tomiyama, K., Doke, N., Murai, A., Katsui, N. and Masamune, T. (1978) *Phytopathology* **68**, 720.
11. Lisker, N. and Kuć, J. (1977) *Phytopathology* **66**, 1356.